TWO COUMARINS FROM THE AEREAL PARTS OF AMYRIS MADRENSIS

X. A. Domínguez, G. Cano, Irene Luna and Antonieta Dieck Departmento de Química, I.T.E.S.M. Monterrey, N.L., México

Key Word Index—Amyris madrensis; Rutaceae; coumarins; dehydrogeijerin; hopeyhopin; triterpene; lupenone; sitosterol.

As part of our research [1] on Mexican Rutaceae, we have studied Amyris madrensis Watt, a shrub common in northeastern Mexico. The aereal parts have been used occasionally as folk medicine, but there are no chemical reports on this plant. The petrol ether extract of dried stems and leaves of A. madrensis on chromatography yielded the triterpene lupenone, sitosterol, the rare coumarin dehydrogeijerin (1) and the new one, hopeyhopin (2). Lupenone was identified by its physical properties and conversion to lupeol.

$$d, 2.25(J = 1) \overset{m, 6.65}{H} \overset{s, 7.21}{H} \overset{d, 6.30}{H} \overset{(J = 9)}{H} \overset{d, 7.68}{H} \overset{(J = 9)}{H} \overset{d, 7.68}{H} \overset{(J = 9)}{H} \overset{d, 7.68}{H} \overset{(J = 9)}{H} \overset{(J = 9)}{H}$$

Dehydrogeijerin (1), mp 129° (M⁺ 258), has been previously isolated only in Geijera parviflora [2]; this coumarin was identified by its mp and spectroscopic properties. The crystals of hopeyhopin 2 C₁₅H₁₄O₅, mp 200-201°, M+ 274, are optical active and fluorescent. The NMR spectrum corresponding to a coumarin [3, 4]. It showed the presence of a methoxyl group, which on its UV and biogenetic grounds was positioned at C-7 and the five carbon epoxy ketone was positioned at C-6. Further, there are only two aromatic singlet protons (δ 8.05, 6.95) which could only be in the 5- and 8-positions. The MS was easily rationalized for the expected fragmentation pattern of such coumarins [5]. On m-chloroperbenzoic acid treatment of 1 a compound was obtained. which showed the same IR, NMR and TLC characteristics as 2 (hopeyhopin).

EXPERIMENTAL

Mp's are uncorr. The 60-MHz NMR spectra were recorded in a Varian A60-A in CDCl₃ with TMS (δ 0.00) as internal standard. IR in KBr, optical activity in CHCl₃.

Isolation of lupenone, sitosterol and the coumarins. Dried powdered stems and leaves (350 g) of A. madrensis (Voucher specimen 7420), were extracted in a Soxhlet with petrol (30-60°). The greenish brown residue (13 g) left after removal of petrol was refluxed 1 hr with 300 ml MeOH [6]. The MeOH soluble portion was evaporated to dryness, the 10 g brownish residue

was chromatographed on Si gel. Elution with C_6H_6 with increasing proportions of MeOH afforded successively: lupenone $C_{30}H_{48}O$, M⁺ 424 (23 mg, mp 160°), IR, NMR, $[\alpha]$, MS, on reduction of lupenone with NaBH₄ lupeol was identified; sitosterol (87 mg, mp 137°), IR, NMR, $[\alpha]$, coTLC and acetate; dehydrogeijerin (1) $C_{15}H_{14}O_4$, M⁺ 258 (178 mg, mp 129°), IR $v_{\rm max}$ KBr cm⁻¹, 3030, 2920, 1724, 1670, 1600, 1480, 1430, 1375, 1370, 1330, 1270, 1200, 1150, 1140, 990, 905, 805, 765, 745. MS: m/e 258 (100%), 257 (90), 243 (M-15) (23), 230 (1.8), 227 (49), 215 (3.6), 213 (9), 203 (40), 189 (7.2), 176 (15), 175 (21), 161 (9), 150 (5.4), 136 (5.4), 118 (7), 96 (6), 88 (9), 82 (24), 68 (13), 56 (16),

s, 1.58
$$\stackrel{s, 4.05}{\text{Me}}$$
 $\stackrel{s, 8.05}{\text{M}}$ $\stackrel{s, 8.05}{\text{H}}$ $\stackrel{d, 6.33}{\text{H}}$ $\stackrel{d}{\text{H}}$ $\stackrel{d, 7.75}{\text{H}}$ $\stackrel{d, 7.75}{\text{H}}$

43 (13), 41 (15). Treatment of 1 with m-chloroperbenzoic acid in CHCl₃ gave crystals of a racemate with IR and NMR spectra identical with the corresponding spectra of hopeyhopine 2.

in CHCl₃ gave crystais of a facemate with 1K and 1NMK spectra identical with the corresponding spectra of hopeyhopine 2. Hopeyhopin 2. $C_{1.5}H_{14}O_{5}$ (M⁺ 274) (100 mg) mp 200–201°, in CHCl₃ $[\alpha]_{589}^{26}$ – 3.7°, $[\alpha]_{578}$ – 4.2°, $[\alpha]_{346}$ – 2.2°, $[\alpha]_{436}$ + 58°. UV λ_{max}^{MeOH} nm (ϵ), 202 (34837), 256 nm (4892), 308 (2266), 331 (2544). IR, ν_{max} KBr cm⁻¹ 3033, 2925, 2875, 1780, 1725, 1650, 1500, 1475, 1450, 1420, 1400, 1360, 1280, 1200, 1160, 1140, 1110, 1010, 905, 895, 815, 805, 735. MS: m/e 274 (19), 259 (5), 245.5*, 216 (2.0), 203 (100), 186 (5), 175 (7), 160 (11), 144 (3), 131 (3), 119 (3), 115.5* (2), 104 (3), 89 (8), 76 (6), 71 (2), 69 (5).

Acknowledgements—We thank Profs George Waller from Oklahoma University and Ernest Eliel from North Carolina University for MS, to Guillermina Vázquez, Angeles Zamudio and E. Ma. Domínguez, for technical assistance. This work was supported by a Syntex, S. A. and Conacyt research grants.

REFERENCES

- Domínguez, X. A., Villegas, D., Rodríguez, V. M. and Zamora, G. (1976) Rev. Latinoamer. Quím. 7, 45.
- Lahey, F. N. and MacLeod, J. K. (1967) Australian J. Chem. 20, 1943.
- 3. Nielsen, B. E. (1970) Dansk. Tiddsskr. 44, 111.
- 4. Steck, W. and Mazurek, M. (1972) Lloydia 35, 418.
- Masrani, K. V., Rama, H. S. and Bafna, S. L. (1974) J. Appl. Chem. Biotechnol. 24, 331.
- Domínguez, X. A. (1973) Métodos de Investigación Fitoquímica. Limusa-Wiley, México.